

NEW β -ORCINOL DEPSIDONES FROM *XANTHOPARMELIA QUINTARIA* AND A *THELOTREMA* SPECIES

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(Received 30 November 1977)

Key Word Index—*Thelotrema* sp.; *Xanthoparmelia quintaria*, Thelotremataceae; lichens; hypostictic acid; hyposalazinic acid; β -orcinol depsidones

Abstract—A new depsidone isolated from a new species of *Thelotrema* was identified as hypostictic acid. *Xanthoparmelia quintaria* contains hypostictic and hyposalazinic acids.

INTRODUCTION

The large crustose lichen family Thelotremataceae, comprising the genera *Leptotrema*, *Ocellularia*, *Phaeotrema* and *Thelotrema*, is widely distributed in tropical and subtropical zones. Although the chemistry of the family has been little studied, it appears to be dominated by the β -orcinol depsidones with the exclusion of the depsides and orcinol compounds [1]. An interesting feature is the joint occurrence of the rare compounds, hypoprotocetraric (1) and 2-*O*-demethylnotatic acids, in many of the species examined [2]. As has been previously proposed by Culberson and Hale [2], these compounds are probably derived from the corresponding depsides, 4-*O*-demethylbarbatic acid and obtusatic acid respectively, which are found in many lichen genera. It is noteworthy, however, that while the aldehydic β -orcinol depsidones (taxonomist's P + compounds) are among the most common of the lichen acids, their corresponding depside precursors are unknown thus suggesting a different mode of formation for these compounds. It seems more likely that the β -orcinol depsidones all arise from a common precursor, hypoprotocetraric acid, by successive oxidations and, in the case of psoromic acid, concurrent decarboxylation (Scheme 1). In this paper we report the isolation of hypostictic acid (4), a probable intermediate in this pathway, from a new species of *Thelotrema*. Its occurrence along with hyposalazinic acid (3) in *Xanthoparmelia quintaria* (Hale) Hale is also reported. The isolation of other intermediates in this biosynthetic pathway will be reported in subsequent papers.

RESULTS AND DISCUSSION

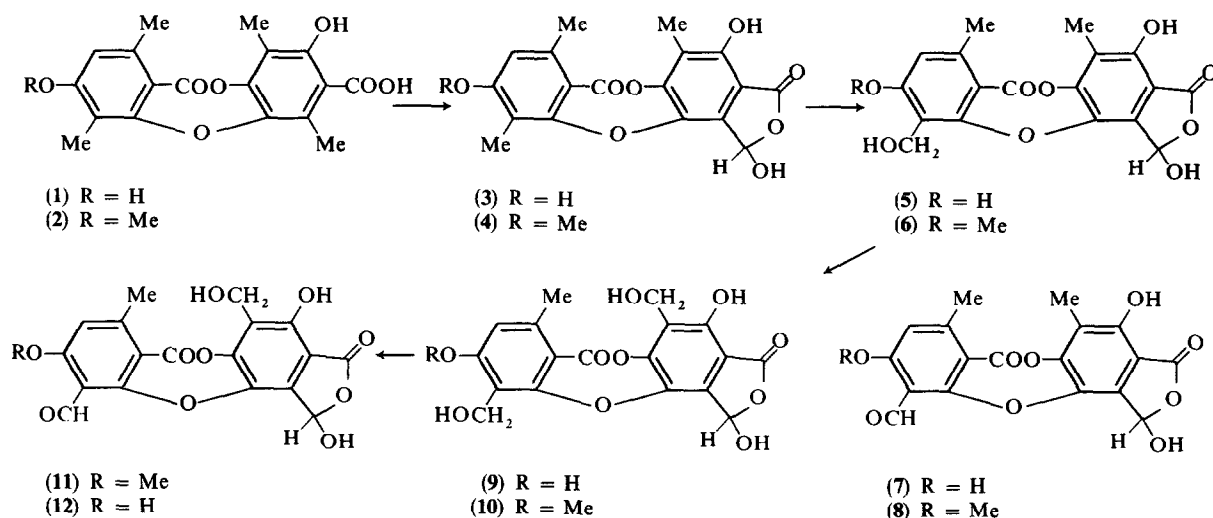
TLC analysis of a micro extract of the lichen, *Thelotrema* sp., showed the presence of two components. Continuous extraction of the lichen with petrol removed some triterpenoid material present in the adhering bark while subsequent extraction with Me₂CO afforded a mixture of two compounds. Compound (4) was readily isolated by fractional crystallization of the mixture from Me₂CO but an insufficient quantity of the minor compound was obtained to permit detailed study.

Compound (4) whose elemental analysis suggested a molecular formula of C₁₉H₁₆O₈ forms a diacetate on treatment with Py/Ac₂O. The IR spectrum shows a sharp peak at 3410 cm⁻¹ (OH) and strong absorptions at 1750 (lactone), 1710 (depsidone), 1600 and 1560 (aromatic). The NMR spectrum (DMSO-d₆) shows two

one-proton signals at δ 6.70 and δ 6.85. The latter signal is considerably sharpened on the addition of D₂O suggesting that the proton is geminal to an OH. This was further confirmed by the large paramagnetic shift produced both on acetylation ($\Delta \sim 0.6$, the signal is also considerably sharpened) and on changing the solvent to Py-d₅ ($\Delta \sim 0.6$). These observations and the high-field position of the signal are characteristic of the proton on the hydroxy lactone ring of the depsidones stictic (8), norstictic (7), salazinic (12), and constictic acids (11) and indicated the presence of this moiety in (4). The solubility of the compound in KHCO₃, although there is no evidence of a carboxylic group in the molecule, supports this hypothesis. Acidification of the basic solution leads to the recovery of (4) unchanged.

Reduction of (4) with Pd/C gives (2), lacking the lactone ring as shown by IR and the replacement of the signal at δ 6.85 by an ArMe signal at δ 2.59. A methoxy (δ 3.87), two relatively low-field Ar-Me resonances (δ 2.20 and δ 2.27), and one at higher field (δ 2.43, typical of a Me ortho to a C=O) complete the spectrum of (4). Double resonance established an ortho relationship between the high-field ArMe and the proton at δ 6.70, while the large aromatic solvent induced shift of the OMe in the spectrum of the diacetate showed this proton also to have an ortho disposition to the OMe. A similar ASIS has been reported [3] in the spectra of stictic tetraacetate and constictic pentaacetate. The substitution pattern on the 'A' ring must therefore be Me, H, OMe, Me as occurs in 4-*O*-methylprotocetraric acid. A second possibility (Me, H, OMe, OH) was considered unlikely on biosynthetic grounds. These data were compatible with the proposed structure (4), hypostictic acid.

Hypostictic acid has not been previously reported as a natural product but was first prepared by Asahina [4] by reduction of naturally occurring stictic acid with Pd/C prepared *in situ* as catalyst. Repeating this procedure we obtained hypostictic acid in high yield, which was identical (IR, NMR, TLC) with the natural product and thus confirmed the structure of (4). We found, however, that commercially available Pd/C catalyst was too active and that its use led to considerable hydrogenolysis of the hydroxylactone ring affording mixtures of (4) and (2). Similarly, (2) was found to be identical with 4-*O*-Mehypoprotocetraric acid formed on reduction of stictic acid under more vigorous conditions. The possibility that the minor component of the lichen might be hyposalazinic acid (3), the 4-*O*-demethylated derivative of (4), was also considered. However, cochromatography of the



extract with authentic (3) prepared by reduction of (12), showed the two compounds to be distinct.

Four unidentified compounds have been previously detected in *Xanthoparmelia quintaria* [5] and have been named PQ1, PQ2, PQ3 and PQ4 by Culberson [6]. Hale also reported the presence of these compounds in *Pseudoparmelia neoquintaria* Hale, in *Relicina abstrusa* (Vaino) Hale, which also produces usnic and norstictic acids, and in some of the brown Parmeliae [7]. On TLC plates these compounds give a red colour, distinct from that of any of the known depsidones, after spraying with H_2SO_4 reagent. Hale, who identified our *Thelotrema* sp., suggested that PQ1 was identical with our hypostictic acid. Cochromatography, in the three standard solvent systems [6], of an acetone extract of *X. quintaria* with hypostictic acid supported this hypothesis. Likewise PQ2 was found to be identical with hyposalazinic prepared by reduction of salazinic acid (12) with Pd/C. Further confirmation was obtained by cochromatography of the acetylation product of the micro extract with the acetate of (4) and hyposalazinic triacetate.

EXPERIMENTAL

Plant material. The *Thelotrema* sp. which was shown to be a new species, was collected in the cloud forest (alt 2300 m) San Eusebio, State of Mérida. A specimen is deposited in the herbarium of this faculty (MFK 20001). A fragment of *Xanthoparmelia quintaria*, a South African endemic, was kindly provided by Dr. M. E. Hale.

Diacetate of (4). Hypostictic acid (200 mg) was dissolved in a mixture of Py and Ac_2O , and after standing at room temp. for ca 18 hr was worked up in the usual manner to give the diacetate (190 mg) mp 244° . IR (KBr) bands at 1785, 1768, 1735, 1360, 1180 and 988 cm^{-1} . PMR (CDCl_3 , δ); 2.15; 2.21, 2.30, 2.41 and 2.50 (3H each, all s, $5 \times \text{Me}$), 3.90 (3H, s, OMe), 6.61 (1H, s, ArH), 7.43 (1H, s, lactone H). (Found; C, 60.48; H, 4.34. $\text{C}_{23}\text{H}_{20}\text{O}_{10}$ requires; C, 60.56; H, 4.38 %).

Synthesis of (4). A soln of PdCl_2 (0.5 %, 8 ml) mixed with activated C (200 mg) and HOAc (8 ml) was reduced at room temp. and press. with H_2 . Stictic acid (100 mg) was then added to the mixture and shaken under H_2 for 12 hr. The reaction mixture was filtered and the C extracted with boiling Me_2CO to afford hypostictic acid identical (mp, IR, PMR) with the natural product, mp 260° d ($\text{Me}_2\text{CO}/\text{H}_2\text{O}$, lit. [4] 264° d).

Hyposalazinic acid (3). Salazinic acid (200 mg) was reduced as in the case of stictic to afford hyposalazinic acid mp 280° d ($\text{Me}_2\text{CO}/\text{H}_2\text{O}$). IR bands at 1700 (br), 1612, 1582, 1264, 1129 and 840 cm^{-1} . PMR ($\text{DMSO}-d_6$, δ); 2.20, 2.27 and 2.33 (3H each, all s, ArMe), 6.63 (1H, s, ArH), 6.66 (1H, s, lactone H).

Triacetate of (4). The acid (100 mg) was suspended in HOAc (4 ml), cooled to 0° and treated with 1 drop of conc H_2SO_4 . After standing ca 18 hr the resulting clear soln was worked up in the normal manner to afford the triacetate of (4) mp $203\text{--}205^\circ$ ($\text{Me}_2\text{CO}/\text{H}_2\text{O}$). IR bands at 1780–1730, 1360, 1190, 1110 and 990 cm^{-1} . PMR (CDCl_3 , δ); 2.13, 2.20, 2.30, 2.32, 2.40 and 2.47 (3H each, all s, $6 \times \text{Me}$), 6.87 (1H, s, ArH), 7.42 (1H, s, lactone H). (Found; C, 59.25; H, 4.05. $\text{C}_{24}\text{H}_{20}\text{O}_{11}$ requires; C, 59.50, H, 4.13 %).

Acknowledgements—The author is indebted to Dr Mason Hale of the Smithsonian Institution Washington, Washington D.C. who identified the lichen and to Sr Daniel Salerno for technical assistance. The financial support of the Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICIT) is gratefully acknowledged.

REFERENCES

1. Culberson, C. F. and Culberson, W. L. (1968) *J. Japan Botany* **43**, 316.
2. Culberson, C. F. and Hale, M. E. (1973) *The Bryologist* **76**, 77.
3. Paolini, J. E. (1972) Tesis de Grado, Universidad de Los Andes, Mérida.
4. Asahina, Y., Yanagita, M. and Omaki, T. (1933) *Chem. Ber.* **66**, 943.
5. Hale, M. E. (1971) *Bot. Notiser* **124**, 343.
6. Culberson, C. F. (1972) *J. Chromatogr.* **72**, 113.
7. Hale, M. E. (1976) *Smithsonian Contrib. Botany* **31**, 39.